ANTITUMOR ACTIVITY OF TRIOXACARCIN C

KAZUHISA FUJIMOTO and MAKOTO MORIMOTO

Pharmaceuticals Research Laboratory, Kyowa Hakko Kogyo Co., Ltd., Shimotogari 1188, Nagaizumi-cho, Suntoh-gun, Shizuoka-ken, Japan

(Received for publication February 19, 1983)

The novel antitumor antibiotic, trioxacarcin C, was studied for antitumor activities against murine tumor systems. When mice with i.p.-inoculated B16 melanoma were given intraperitoneal injections of trioxacarcin C, the maximal T/C% was 164 by successive administration of 0.125 mg/kg/day (day 1~10). It also gave the prolongation of life span of mice bearing i.p. P388 leukemia (T/C 141%) by i.p. injection for 10 days, and inhibited the growth of sarcoma 180 (T/C 42%) and Lewis lung carcinoma implanted s.c. (T/C 23%) by i.v. administration for 6 or 7 days. It inhibited the growth of P388 leukemia cells *in vitro* and showed significant inhibition on the colony formation of HeLa S₈ cells. DNA and RNA synthesis were more strongly inhibited than protein synthesis by trioxacarcin C. Also, it induced strand scission of PM-2 DNA without reducing agents or metals. It did not effect the number of white blood cells and blood urea nitrogen value of the peripheral blood.

Trioxacarcin, a group of antibiotics with novel polycyclic chromophores¹⁾, was isolated from the culture broth of *Streptomyces bottropensis* DO-45.^{1,2)} This antibiotic is active against Gram-positive bacteria such as *Staphylococcus aureus* and *Bacillus subtilis*, but is weakly active against Gram-negative bacteria such as *Escherichia coli* and *Klebsiella pneumoniae*. The preliminary results of antitumor activity of trioxacarcin against sarcoma 180 and P388 leukemia were described in the previous report,¹⁾ but the characterization of trioxacarcin C as an anticancer agent has not been previously elucidated.

This paper describes the antitumor activity against various murine tumors and cell growth inhibition of trioxacarcin C. The mechanism of action of trioxacarcin C was studied on the macromolecular synthesis of P388 leukemia cells *in vitro* and PM-2 DNA-cleaving activity. Myelosuppression and nephrotoxicity were also studied in mice.

Materials and Methods

Chemicals

Trioxacarcin C was obtained from Dr. F. TOMITA of Tokyo Research Laboratory of Kyowa Hakko. Mitomycin C (Kyowa Hakko Kogyo Co., Ltd., Japan), bleomycin (Nippon Kayaku Co., Ltd., Japan) and neocarzinostatin (Yamanouchi Seiyaku Co., Ltd., Japan) were used reference antitumor agents. [6-³H]Thymidine, [5-³H]uridine and L-[4,5-³H]leucine were purchased from the Radiochemical Centre Amersham (England). PM-2 phage DNA was purchased from the Boehringer Manheim Yamanouchi Co., Ltd. (Tokyo, Japan)

Animals

Male ddY (18 ~ 20 g weight), CDF₁ (5 ~ 8 weeks old) and BDF₁ mice (5 ~ 8 weeks old) were purchased from the Shizuoka Agricultural Co-operative Association for Laboratory Animals (Hamamatsu, Japan).

Tumors and Toxicity

Antitumor activity and toxicity were measured and calculated as described previously.⁸⁾

 LD_{50} was calculated by BEHRENS-KÖRBER method from the number of survivors at 30 days after a single intravenous or intraperitoneal administration of drug into ddY mice.

THE JOURNAL OF ANTIBIOTICS

Peripheral white blood cells and blood urea nitrogen were determined according to the method described previously.³⁾ Differential counts of leucocytes were made on Giemsa stained smears of pretreatment blood and posttreatment blood on day 4, 7, 10 and 15.

Culture of Leukemia P388 Cells

Leukemia P388 cells were cultured in RPMI 1640 (Gibco Laboratories, New York, U.S.A.) supplemented with 5% fetal bovine serum (Gibco Laboratories), 50 μ M of 2-mercaptoethanol, 100 U/ml of benzylpenicillin (Meiji Seika Co., Ltd., Tokyo, Japan) and 100 μ g/ml of streptomycin (Kyowa Hakko Co., Ltd., Tokyo, Japan). For testing the growth inhibitory activity of drugs, 2×10⁴ cells precultured for 24 hours at 37°C in a humidified atmosphere of 5% CO₂ in air, were treated with drugs for 1 hour (pulse exposure) or for 72 hours (continuous exposure). The cell count were made at 72 hours after addition of drug.

Incorporation of the Labeled Precursors into Macromolecule

Logarithmically growing P388 cells $(1 \times 10^5/\text{ml})$ were cultured with trioxacarcin C. [^aH]Thymidine (103 mCi/mg), [^aH]uridine (114 mCi/mg) or [^aH]leucine (385 mCi/mg) were added at 1, 3 and 5 hours after addition of trioxacarcin C. The cells collected on membrane filter GF/F (Whatman Co., Ltd., England) were washed with cold phosphate buffered saline (pH 7.4), and rinsed with cold 5% trichloro-acetic acid several times. The radioactivity on the filter was counted by a liquid scintillation counter (Aloka LSC-653).

Culture of HeLa S₃ Cells

HeLa S₃ cells were cultured in Eagle minimum essential medium (MEM, Nissui Seiyaku Co., Ltd., Tokyo, Japan) supplemented with 10% fetal bovine serum, 50 μ g/ml of kanamycin and 292 μ g/ml of L-glutamine.

For testing the lethal effect of drugs, 150 or 750 HeLa S_s cells were seeded in plastic Petri dishes (60 × 15 mm, Falcon, California, U.S.A.). After incubating for 3 hours at 37°C in a humidified atmosphere of 5% CO₂ in air, the cells were treated with drugs for 1 hour, rinsed with phosphate balanced saline (pH 7.4), and fed the fresh prewarmed medium. The cells were cultured for 10~12 days for colony development, and fixed and stained with 4% Giemsa solution for counting the number of colonies. The plating efficiency of control cells were 70~90% in all experiments.

Analysis of DNA Degradation in Agarose Gel Electrophoresis

The reaction mixtures contained 0.5 μ g PM-2 DNA, 40 mM Tris-HCl buffer (pH 7.8) and various concentration of drug. After incubation for 30 minutes at room temperature, the reaction was terminated by addition of 5 μ l of 50 mM EDTA. After addition of 0.1% bromophenol blue in 75% glycerol, the resultant mixture was electrophoresed on 0.8% agarose gel according to the procedure described by SUZUKI *et al.*⁴⁾ After electrophoresis, the gels were stained in electrophoresis buffer containing 0.5 μ g/ml of ethidium bromide.

Results

Acute Toxicity

The LD_{50} of trioxacarcin C was about 1.0 mg/kg by a single intraperitoneal (i.p.) or intravenous (i.v.) administration in ddY mice.

Antitumor Activity

Summary of the antitumor activity of trioxacarcin C against murine tumors is shown in Table 1. Trioxacarcin C inhibited the growth of solid sarcoma 180 by successive i.v. injection with minimal T/C% of 42 on day 7. But a single injection gave only 46% inhibition on tumor growth. The growth of Lewis lung carcinoma was inhibited by the successive i.v. treatment of trioxacarcin C with minimal T/C% of 23 on day 13. Maximal T/C% of 121 for survival day was obtained in this schedule, but prolongation of life span was not significant.

Tumor		Drug		Dose	TIC
System	Site	Route	Schedule	(mg/kg/day)	T/C (%)
Sarcoma 180	S.C.	i.v.	Day 1	0.75	54
	s.c.	i.v.	Days $1 \sim 6$	0.5	42
P388	i.p.	i.p.	Day 1	0.25	130
	i.p.	· i.p.	Days 1, 5, 9	0.125	136
	i.p.	i.p.	Days 1~10	0.125	141
B16	i.p.	i.p.	Day 1	0.25	149
	i.p.	i.p.	Days 1~10	0.125	164
3LL	s.c.	i.v.	Day 1	0.9	67
	S.C.	i.v.	Days 1, 7, 13	0.4	74
	s.c.	i.v.	Days $1 \sim 7$	0.3	23

Table 1. Effect of trioxacarcin C on murine tumor.

Trioxacarcin C prolonged the life span of leukemia P388 bearing mice. Almost equal T/ C%s were observed with three schedules examined (single, intermittent and successive). A significant activity was shown in i.p.-inoculated B16 melanoma. Maximal effect (T/C 164%) was obtained in mice treated with successive i.p. injection of 0.125 mg/kg/day trioxacarcin C.

Effect on the Growth of Leukemia

P388 In Vitro

The growth of leukemia P388 cells was markedly inhibited by trioxacarcin C in vitro. As

Table 2. Effect of trioxacarcin C on the growth of P388 leukemia cell.

	IC_{50}		
Drug	Continuous exposure (72 hours) (MM)	Pulse exposure (1 hour) (mM)	
Trioxacarcin C	8.24×10 ⁻⁶	1.39×10 ⁻⁴	
Bleomycin	1.07×10^{-2}	2.24×10^{-2}	
Neocarzinostatin	4.92×10 ⁻⁵	3.44×10 ⁻⁶	

Exponentially growing P388 leukemia cells were treated with each drug for 1 or 72 hours. Count of cells was made at 72 hours after addition of drugs.

shown in Table 2, the IC₁₀ (concentration which gave 50% cell number of control) was 8.24×10^{-6} mM at continuous exposure, and 1.39×10^{-4} mM at pulse exposure. The IC₁₀ value of trioxacarcin C was more than 100 times lower than that of bleomycin by pulse and continuous exposure. Neocarzinostatin was more inhibitory than trioxacarcin C by pulse exposure.

Lethal Effect on HeLa S3 Cells

The dose-survival curve of HeLa S_s cells exposed to trioxacarcin C for 1 hour is presented in Fig. 1. Trioxacarcin C-treated cells showed a dose-dependent exponential survival curve with a slight shoulder at low concentration. The mean lethal dose (D_0 , the dose necessary to give 37% survival) was 1.30×10^{-5} mM, and n (extrapolating value of an exponential portion of dose survival curve) was 1.60 for trioxacarcin C. For bleomycin-treated cells, the D_0 and n values were 1.46×10^{-2} mM and 0.69, and for neocarzinostatin, 1.32×10^{-6} mM and 3.48, respectively.

Effect on Macromolecular Synthesis

Fig. 2 shows the inhibitory effect of trioxacarcin C on the incorporation of $^{\circ}$ H-labelled precursors into macromolecules of leukemia P388 cells. At a concentration of 1.39×10^{-4} mM (IC₅₀ at pulse exposure), trioxacarcin C suppressed DNA and RNA synthesis more strongly than protein synthesis (time

Fig. 1. Lethal effect of trioxacarcin C on HeLa $S_{\scriptscriptstyle 3}$ cells.

HeLa S_{\circ} cells was exposed to each drug for 1 hour. Each point represents mean value of 3 experiments with trioxacarcin C.

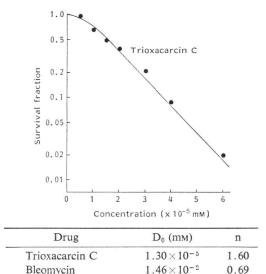


Fig. 3. Agarose gel electrophoresis of PM-2 DNA following incubation with trioxacarcin C.

 1.32×10^{-6}

3.48

Neocarzinostatin

1 and 8: Control, PM-2 DNA, 2 to 7: trioxacarcin C, 4 μ g/ml, 10 μ g/ml, 20 μ g/ml, 40 μ g/ml, 100 μ g/ml, 200 μ g/ml.

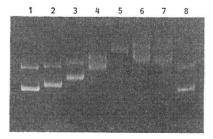
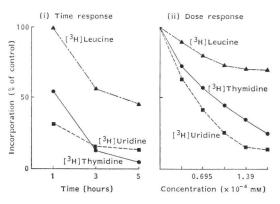


Fig. 2. Inhibitory effect of trioxacarcin C on incorporation of labelled precursors into P388 primary culture cells.

(i) The labelled precursors were added at the indicated time after addition of trioxacarcin C and radioactivity was counted after pulse labelling for 1 hour.

(ii) The labelled precursors were added at 3 hours after addition of trioxacarcin C and radioactivity was counted after pulse labelling for 1 hour.



response). From the dose response study, there was no difference on the inhibition of incorporation of [⁸H]thymidine and [⁸H]uridine into leukemia P388 cells.

DNA Degradation

Covalently closed circular PM-2 DNA was treated with various concentrations of trioxacarcin C. According to AAJI and BORST,⁵⁾ the fastest moving zone corresponds to covalently closed circular DNA (form I), the most slowly moving one to the open circular DNA (form II), and the intermediate to the linear DNA. As

shown in Fig. 3, 100 μ g/ml of trioxacarcin C caused single strand scission of PM-2 DNA as well as change in the mobility (lane 6). As shown in the lane 7, the amount of the linear DNA was increased, while that of the open circular DNA was decreased. From these results, trioxacarcin C could make double strand scission of PM-2 DNA.

Bone Marrow Toxicity and Renal Toxicity

Fig. 4 shows the time course change of white blood cells and blood urea nitrogen in mice after i.v. treatment of trioxacarcin C and mitomycin C (MMC). Trioxacarcin C exhibited no significant decrease in white blood cells but slight increase at a single dose of 1.2 mg/kg with concomitant increase in neutrophiles (data not shown). Mitomycin C gave significant decrease in white blood cells with concomitant decrease in lymphocytes.

The blood urea nitrogen value of mice treated with trioxacarcin C did not deviate from that of

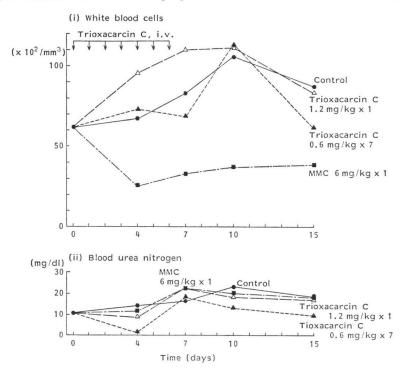


Fig. 4. Effect of trioxacarcin C on peripheral white blood cells and blood urea nitrogen.

control mice.

Discussion

Trioxacarcin C gave a prolongation of the life span of mice bearing i.p.-inoculated P388 leukemia and B16 melanoma by a single and successive administrations.

For solid tumors, a single administration of trioxacarcin C not inhibit the growth of solid sarcoma 180 and Lewis lung carcinoma. The successive injection (day $1 \sim 6$) gave T/C% of 42 against sarcoma 180 on day 7 and all animals treated with 0.5 mg/kg/day survived on day 7. But, as mortality of treated animal was not observed thereafter, we could not conclude the effectiveness of trioxacarcin C against sarcoma 180. For solid tumor of Lewis lung carcinoma, 7 successive i.v. administrations of trioxacarcin C inhibited the growth of tumor and gave a slight prolongation of life span with T/C% of 121, showing no sign of toxic death. In the previous report, LD₅₀ was reported to be 2 mg/kg by a single i.p. administration,¹⁾ but further studies with purified sample gave 1 mg/kg for LD₅₀ by a single injection. The discrepancy of the values between previous and this report might result from the purity of sample.

Trioxacarcin C may interact with DNA and give DNA strand scission. But the mechanism of trioxacarcin C might be somewhat different from that of bleomycin,⁶⁾ neocarzinostatin⁷⁾ and macromomycin,⁴⁾ because reducing agents were required for DNA strand scission by latter compounds. Trioxacarcin C-induced DNA breakage might contribute to its cytotoxicity. Further study on DNA breaking activity of trioxacarcin C will be performed.

Because of DNA strand scission activity of trioxacarcin C, the effect of trioxacarcin C on the *in vitro* cell growth was compared with those of bleomycin or neocarzinostatin by assays of P388 leukemia cells with suspension culture and HeLa S_{\circ} cells with colony formation. Trioxacarcin C was about 100 and 10 times more potent than bleomycin and mitomycin C,^{\$} respectively, against the growth of P388 leukemia cells *in vitro* with pulse exposure. It was also more active than bleomycin for the colony formation.

ing ability of HeLa S_3 cells. A slight shoulder on the dose-survival curve of HeLa S_3 cells might suggest that trioxacarcin C induced DNA repair synthesis from sublethal damage.

Trioxacarcin C was reported to inhibit DNA synthesis without significant effect on RNA and protein synthesis in *Bacillus subtilis*,²⁾ but, there is no difference on the inhibition % of DNA and RNA synthesis at various periods after addition and the same concentration of drugs in P388 leukemia cells. Trioxacarcin C might inhibit different steps of macromolecular synthesis between mammalian cells and microorganisms.

As the number of peripheral white blood cells did not decrease by a single and successive injection of trioxacarcin C, the bone marrow toxicity of trioxacarcin C might not be significant in mice. But the increase in neutrophiles count might suggest the irritating activities of trioxacarcin C.

While the antitumor activity and spectrum of trioxacarcin C was not superior to mitomycin C or adriamycin, the lower myelosuppressive activity of trioxacarcin C as compared with mitomycin C or adriamycin, might be one of the interesting characteristics for further studies of this compound.

Acknowledgment

The authors thank Miss M. WADA for the technical assistance with the experiment and Miss E. NANJO for typing the manuscript.

References

- TOMITA, F.; T. TAMAOKI, M. MORIMOTO & K. FUJIMOTO: Trioxacarcins, novel antitumor antibiotics. I. Producing organisms, fermentation and biological activities. J. Antibiotics 34: 1519~1524, 1981
- TAMAOKI, T.; K. SHIRAHATA, T. IIDA & F. TOMITA: Trioxacarcins, novel antitumor antibiotics. II. Isolation, physicochemical properties and mode of action. J. Antibiotics 34: 1525~1530, 1981
- MORIMOTO, M.; M. FUKUI, S. OHKUBO, T. TAMAOKI & F. TOMITA: Tetrocarcins, new antitumor antibiotics.
 Antitumor activity of tetrocarcin A. J. Antibiotics 35: 1033~1037, 1982
- SUZUKI, H.; K. MIURA & N. TANAKA: DNA-cleaving potentials of macromomycin and auromomycin: A comparative study. Biochem. Biophys. Res. Comm. 89: 1281 ~ 1286, 1979
- 5) AAJI, C. & P. BORST: The gel electrophoresis of DNA. Biochem. Biophys. Acta 269: 192~200, 1972
- SUZUKI, H.; K. NAGAI, H. YAMAKI, N. TANAKA & H. UMEZAWA: On the mechanism of action of bleomycin: Scission of DNA strands *in vitro* and *in vivo*. J. Antibiotics 22: 446~448, 1969
- ISHIDA, R. & T. TAKAHASHI: Role of mercaptoethanol in *in vitro* DNA degradation by neocarzinostatin. Cancer Res. 38: 2617~2620, 1978